

HIGHLY OXYGENATED FUROEREMOPHILANE DERIVATIVES FROM *SENECIO ZOELLNERI*

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Abstract—The aerial parts of *Senecio zoellneri* afforded, in addition to known compounds, five oxygenated furoeremophilane derivatives and three *seco*-furoeremophilanolides, all of which are probably formed by oxidation of 3 β -hydroxy-6 β -acyloxyureyopsin.

INTRODUCTION

Many species from the large genus *Senecio* have been investigated chemically. The most widespread compounds are furoeremophilanes and their derivatives. We have now studied the chemistry of *S. zoellneri* Mart. et Quez.

RESULTS AND DISCUSSION

The aerial parts of *S. zoellneri* afforded spathulenol, *p*-hydroxyacetophenone, tremetone, cinnamyl angelate, the furoeremophilane 10 and the highly oxygenated derivatives 2–9.

The structure of cinnamyl angelate followed directly from its ¹H NMR spectrum. The ¹H NMR spectra of 5 and 6 (Table 1) differed only in the signals of the ester residues. Spin decoupling allowed the assignment of all signals. The sequences obtained indicated the presence of eremophilanolides with a 3 β -hydroxy, a 6 β -acyloxy and a 8 α -methoxy group. The configuration at C-8 followed from the chemical shifts of the neighbouring protons which differed from the shifts of similar lactones with opposite configuration at C-8 [1, 2].

The ¹H NMR spectrum of 4 differed from that of 5 by the absence of the methoxy singlet. All data agree with the presence of the corresponding 8 α -hydroxy derivative. The ¹H NMR spectra of 2 and 3 (Table 1) again differed only in the signals of the ester groups. In part the data were close to those of 5 and 6. However, the signals of H-6, H-9 and H-13 differed markedly. The molecular formula of 2 is C₁₉H₂₆O₆ indicating that it is an isomer of 4. The chemical shifts of H-13 in 2 and 3 required a methyl group on an oxygen-bearing carbon and singlets at δ 5.30 and 5.25 agree with the presence of a 7,8,11,12-bis-epoxide. The configuration of the epoxides could not be determined. However, inspection of models and the proposed formation favours the α -orientation. A similar compound derived from a furocadinane has been isolated from a

Baccharis species [3]. Most likely compounds 2 and 3 are formed via the endoperoxide 1, the addition product of the corresponding furoeremophilane and singlet oxygen. The bis-epoxides 2 and 3 then could be transformed to the lactones of type 4 as shown in the Scheme.

The structures of the lactones 7 and 8 could be deduced from their ¹H NMR spectra (Table 1) which were similar to those of the corresponding 3-desacetoxy derivative of 8 [4]. The splitting of several ¹H NMR signals showed that both 7 and 8 are mixtures of C-8 epimers which could not be separated. As the relation was *ca* 1:2, the ¹H NMR signals were easily assigned. The couplings of H-3 indicated the proposed configuration at C-3, while that at C-6 was supported by biogenetic considerations as again these compounds are most likely derived from 4.

The ¹H NMR spectrum of 9 (Table 1) differed markedly from those of 4–8. The presence of a new methylene group followed from the signals at δ 4.86 and 5.23, while signals at δ 4.49 and 4.45 require the presence of two hydroxy groups. In agreement with all data structure 9 was proposed. The stereochemistry was confirmed by NOED [H-14 with H-15 (7%), H-13 (7%) and H-9' (8%); H-13 with H-6 (5%); H-4 with H-3 (4%); OMe with H-8 (10%); H-1 with H-9 (8%) and H-3 with H-6 (8%)]. Lactone 9, which we have named *secosenzoellneride*, is probably formed by fragmentation of the 9-hydroxy derivative of 4.

EXPERIMENTAL

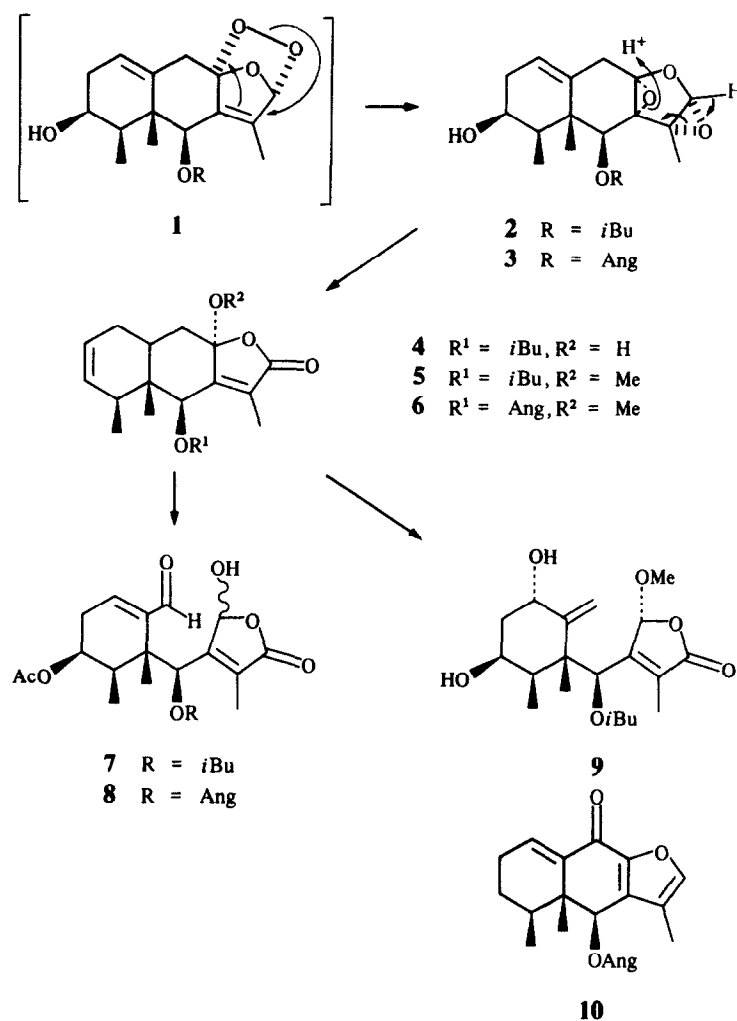
The air-dried plant material (448 g, voucher Niemeyer 8986, deposited in the Herbarium of the University of Chile, Santiago, collected in May 1989 in the Region de Farapaca) was extracted with a mixt. of solvents comprising petrol, Et₂O and MeOH in equal amounts. The extract was freed from solvent, defatted by MeOH and subsequently separated by CC (silica gel) using petrol, Et₂O and MeOH with increasing polarity. The frs obtained from the column were later further separated and purified by repeated TLC and HPLC as reported previously [5] to give 10 mg 2 (HPLC: MeOH–H₂O, 3:2), 2 mg 3 (TLC1), 6 mg 4 (TLC2), 5 mg 5 (TLC2), 3 mg 6 (TLC2), 4 mg 7 (TLC1), 5 mg 8 (TLC1), 3 mg 9 (TLC1), 3 mg cinnamylangelate, 6 mg tremetone,

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Table 1. $^1\text{H NMR}$ spectral data of compounds 2–9 (400 MHz, CDCl_3 , δ -values)

H	2	3	4	5	6	7		8		9
1	5.47 <i>br q</i>	5.50 <i>br q</i>	5.68 <i>m</i>	5.65 <i>ddd</i>	5.66 <i>ddd</i>	6.89	6.85 <i>dd</i>	6.89	6.84 <i>dd</i>	4.45 <i>br t</i>
2	2.53 <i>dddd</i>	2.55 <i>dddd</i>	2.30 <i>br dt</i>	2.30 <i>br dt</i>	2.30 <i>br dt</i>	2.87	2.83 <i>ddd</i>	2.90	2.85 <i>ddd</i>	1.99 <i>br d</i>
2'	2.21 <i>br dt</i>	2.22 <i>br dt</i>	2.15 <i>m</i>	2.16 <i>m</i>	2.16 <i>m</i>	2.40 <i>m</i>		2.40 <i>m</i>		1.73 <i>br dt</i>
3	3.95 <i>br d</i>	3.96 <i>br d</i>	4.08 <i>ddd</i>	4.01 <i>ddd</i>	4.02 <i>ddd</i>	5.42 <i>m</i>		5.45 <i>m</i>		4.47 <i>dt</i>
4	1.77 <i>dq</i>	1.82 <i>dq</i>	1.82 <i>dq</i>	1.85 <i>dq</i>	1.95 <i>m</i>	2.41	2.45 <i>dq</i>	2.40 <i>m</i>		2.28 <i>br q</i>
6	5.25 <i>s</i>	5.30 <i>s</i>	5.68 <i>q</i>	5.40 <i>q</i>	5.47 <i>d</i>	5.78	5.81 <i>s</i>	5.86	5.91 <i>s</i>	6.58 <i>s</i>
8	—	—	—	—	—	5.67 <i>br s</i>		5.69 <i>br s</i>		5.58 <i>br s</i>
9	2.92 <i>dd</i>	2.94 <i>dd</i>	2.74 <i>d</i>	2.78 <i>d</i>	2.79 <i>d</i>	9.29 } 9.36 <i>s</i> }		9.30 } 9.36 <i>s</i> }		5.23 <i>s</i> }
9'	2.38 <i>d</i>	2.42 <i>d</i>	2.50 <i>ddd</i>	2.43 <i>ddd</i>	2.44 <i>ddd</i>					4.86 <i>d</i> }
12	5.30 <i>s</i>	5.35 <i>s</i>	—	—	—	—	—	—	—	—
13	1.64 <i>s</i>	1.64 <i>s</i>	1.88 <i>d</i>	1.92 <i>d</i>	1.85 <i>d</i>	2.19	2.10 <i>s</i>	2.17	2.19 <i>s</i>	2.10 <i>s</i>
14	1.28 <i>s</i>	1.32 <i>s</i>	1.05 <i>s</i>	1.07 <i>s</i>	1.10 <i>s</i>	1.35	1.36 <i>s</i>	1.37	1.38 <i>s</i>	1.10 <i>s</i>
15	1.04 <i>d</i>	1.07 <i>d</i>	1.03 <i>d</i>	1.06 <i>d</i>	1.05 <i>d</i>	0.85	0.88 <i>s</i>	—	—	0.82 <i>d</i>
OCOR	2.62 <i>qq</i>	6.13 <i>qq</i>	2.74 <i>qq</i>	2.72 <i>qq</i>	6.30 <i>qq</i>	2.62 <i>qq</i>		6.16 <i>qq</i>		2.68 <i>qq</i>
	1.23 <i>d</i>	2.04 <i>dq</i>	1.29 <i>d</i>	1.30 <i>d</i>	2.08 <i>dq</i>	1.20	1.18 <i>d</i>	—	1.99 <i>dq</i>	1.25 <i>d</i>
	1.22 <i>d</i>	1.96 <i>dq</i>	1.25 <i>d</i>	1.26 <i>d</i>	2.01 <i>dq</i>	1.19	1.16 <i>d</i>	1.95	1.96 <i>dq</i>	1.20 <i>d</i>
OMe	—	—	—	3.19 <i>s</i>	3.22 <i>s</i>	—	—	—	—	3.43 <i>s</i>

J [Hz]: Compounds 2 and 3: 1,2 = 1,2' = 1,9 ~ 3; 2,2' = 18; 2,3 = 5; 3,4 = 2; 4,15 = 7; 9,9' = 15; compounds 4–6: 1,2 = 2,3 ~ 4.5; 1,2' = 3; 1,9' = 2',9' ~ 2; 2,2' = 18; 2',3 = 9; 3,4 = 2.5; 4,15 = 7; 6,13 = 1; 9,9' = 13; compounds 7 and 8: 1,2 = 3; 1,2' = 4; 2,2' = 20; 2,3 = 6; 3,4 = 2.5; 4,15 = 7; compound 9: 1,2 = 1,2' = 4; 1,9' ~ 1; 2,2' = 2',3 = 12; 2,3 = 3,4 = 3.5; 4,15 = 7; OiBu: 2,3 = 2,4 = 7; OAng: 3,4 = 7; 3,5 = 4,5 = 1.5.



25 mg spathulenol and 50 mg *p*-hydroxyacetophenone. Known compounds were identified by comparing their spectral data with those of the reference compounds. TLC1: C₆H₆-CH₂Cl₂-Et₂O (1:1:2); TLC2: C₆H₆-CH₂Cl₂-Et₂O (3:3:4).

3β-Hydroxy-6β-isobutyryloxy-7,8,11,12-bis-epoxyfuroeremophil-1(10)-ene (2). IR ν_{max}^{CHCl₃} cm⁻¹: 3520 (OH), 1750 (CO₂R); MS *m/z* (rel. int.): 350.173 [M]⁺ (3) (calc. for C₁₉H₂₆O₆: 350.173), 262 [M-RCO₂H]⁺ (46), 205 [262-C₃H₅O]⁺ (90), 120 (77), 71 [RCO]⁺ (100).

3β-Hydroxy-6β-angeloyloxy-7,8,11,12-bis-epoxyfuroeremophil-1(10)-ene (3). IR ν_{max}^{CHCl₃} cm⁻¹: 3600 (OH), 1730 (C=CCO₂R); MS *m/z* (rel. int.): 362.173 [M]⁺ (1) (calc. for C₂₀H₂₆O₆: 362.173), 262 [M-RCO₂H]⁺ (44), 205 (26), 120 (26), 83 [RCO]⁺ (100), 55 [83-CO]⁺ (60).

3β,8α-Dihydroxy-6β-isobutyryloxyeremophil-1(10),7(11)-dien-12,8β-olide (4). IR ν_{max}^{CHCl₃} cm⁻¹: 3420 (OH), 1750 (γ-lactone, CO₂R); MS *m/z* (rel. int.): 350 [M]⁺ (0.5), 332.163 [M-H₂O]⁺ (2) (calc. for C₁₉H₂₄O₅: 332.163), 244 [332-RCO₂H]⁺ (42), 140 (90), 121 (52), 106 (62), 71 [RCO]⁺ (100).

3β-Hydroxy-6β-isobutyryloxy-8α-methoxyeremophil-1(10),7(11)-dien-12,8β-olide (5). IR ν_{max}^{CHCl₃} cm⁻¹: 3600 (OH), 1780 (γ-lactone), 1750 (CO₂R); MS *m/z* (rel. int.): 364.189 [M]⁺ (1) (calc. for C₂₀H₂₈O₆: 364.189), 332 [M-MeOH]⁺ (1.3), 276 [M-RCO₂H]⁺ (23), 244 [276-MeOH]⁺ (17), 121 (26), 71 [RCO]⁺ (100).

3β-Hydroxy-6β-angeloyloxy-8α-methoxyeremophil-1(10),7(11)-dien-12,8β-olide (6). IR ν_{max}^{CHCl₃} cm⁻¹: 3600 (OH), 1775 (γ-lactone), 1720 (C=CCO₂R); MS *m/z* (rel. int.): 376.189 [M]⁺ (2.5) (calc. for C₂₁H₂₈O₆: 376.189), 344 [M-MeOH]⁺ (2.5), 276 [M-RCO₂H]⁺ (24), 244 [276-MeOH]⁺ (18), 121 (35), 107 (42), 83 [RCO]⁺ (100), 55 [83-CO]⁺ (47).

3β-Acetoxy-6β-isobutyryloxy-seco-macrotolide (7). IR ν_{max}^{CCl₄} cm⁻¹: 2720, 1700 (C=CCHO), 1790 (γ-lactone), 1740 (CO₂R); CIMS *m/z* (rel. int.): 279 [M+1-RCO₂H, ketene]⁺ (1.5), 261 [279-H₂O]⁺ (1), 221 (3), 203 (4), 133 (26), 73 (100).

3β-Acetoxy-6β-angeloyloxy-seco-macrotolide (8). IR ν_{max}^{CCl₄}

cm⁻¹: 2720, 1700 (C=CCHO), 1790 (γ-lactone), 1745 (OAc), 1720 (C=CCO₂R); CIMS *m/z* (rel. int.): 343 [M+1-HOAc, H₂O]⁺ (0.5), 279 [M+1-RCO₂H, ketene]⁺ (2.5), 261 [279-H₂O]⁺ (2.5), 203 (100).

seco-Senzoellneride (9). IR ν_{max}^{CHCl₃} cm⁻¹: 3600 (OH), 1750 (γ-lactone, CO₂R); MS *m/z* (rel. int.): 382 [M]⁺ (0.5), 364.189 [M-H₂O]⁺ (1.7) (calc. for C₂₀H₂₈O₆: 364.189), 294 [M-RCO₂H]⁺ (12), 276 [294-H₂O]⁺ (12), 262 [294-MeOH]⁺ (18), 244 [262-H₂O]⁺ (17), 158 (51), 140 (100), 71 [RCO]⁺ (82).

Cinnamylangelate. IR ν_{max}^{CCl₄} cm⁻¹: 1725 (C=CCO₂R); MS *m/z* (rel. int.): 216.115 [M]⁺ (8) (calc. for C₁₄H₁₆O₂: 216.115), 117 [M-OAng]⁺ (40), 83 [RCO]⁺ (100); ¹H NMR (CDCl₃): δ 7.40 (*br d*, H-2, H-6), 7.33 (*br t*, H-3, H-5), 7.25 (*br t*, H-4), 6.18 (*dt*, H-7), 5.84 (*dt*, H-8), 4.81 (*dd*, H-9); OAng: 6.10 *qq*, 2.00 *dq*, 1.92 *dq*; *J* [Hz]: 2,3 = 3,4 ~ 8; 7,8 = 16; 7,9 = 1.5; 8,9 = 7; 3,5 = 4,5 = 1.5.

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REFERENCES

- Dupre, S., Grenz, M., Jakupovic, J., Bohlmann, F. and Niemeyer, H. M. (1990) *Phytochemistry* **29** (in press).
- Merikli, A. H., Merikli, F., Jakupovic, J., Bohlmann, F., Dominguez, X. A. and Vega, H. S. (1989) *Phytochemistry* **28**, 1149.
- Bohlmann, F., Banerjee, S., Jakupovic, J., King, R. M. and Robinson, H. (1984) *Rev. Latinoam. Quim.* **15**, 71.
- Bohlmann, F., Gupta, R. K., Jakupovic, J., King, R. M. and Robinson, H. (1981) *Phytochemistry* **20**, 1155.
- Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1984) *Phytochemistry* **23**, 1979.